Hydroxy fatty acids in fresh snow samples from northern Japan: long-range atmospheric transport of Gram-negative bacteria by Asian winter monsoon

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Abstract. Hydroxy fatty acids (FAs) in fresh snow from Sapporo, one of the heaviest snowfall regions in the world, have been studied to ascertain the airborne bacterial endotoxin concentrations and their biomass. The presence of β-hydroxy FAs (C₉–C₂₈), constituents of the Gram-negative bacterium (GNB), suggests long-range transport of soil microbes. Likewise, the occurrence of α- and ω-hydroxy FAs (C₉–C₃₀ and C₀–C₂₈, respectively) in snow reveals their contribution from epicuticular waxes and soil microorganisms. Estimated endotoxin and GNB mass can aid in assessing their possible impacts on the diversity and functioning of aquatic and terrestrial ecosystems, as well as lethal effects on pedestrians through dispersal of microbes. Air mass back trajectories together with hydroxy FAs reveal their sources from Siberia, the Russian Far East and northern China by the Asian monsoon. This study highlights the role of fresh snow that reduces the human health risk of GNB and endotoxin by the scavenging from air.

1 Introduction

Lipid biomarkers from terrigenous plants, algae, fungi and soil microorganisms have been reported extensively in aerosols (Conte and Weber, 2002; Gagosian et al., 1987, 1981; Kawamura, 1995; Kawamura et al., 2003; Simoneit, 1977; Simoneit et al., 2004), sediments (Kawamura, 1995; Kawamura and Ishiwatari, 1984; Kawamura et al., 1987; Zhang et al., 2014), ice cores (Sankelo et al., 2013) and rain/snow (Kawamura and Kaplan, 1986; Satsumabayashi et al., 2001; Yamamoto et al., 2011). These studies have utilized fatty acids as a proxy to assess the terrigenous contribution of higher plant waxes to various environmental samples owing to their abundant presence in biopolymers of plants and microorganisms. Similarly, certain hydroxy fatty acids (e.g., C₁₀–C₁₈ β-hydroxy FAs) have been proposed as a tracer to understand the airborne bacterial transport (Tyagi et al., 2015).

Among the airborne soil microbes, the Gram-negative bacterium (GNB) is one of most extensively studied bacteria and is documented in aerosols, snow and rain samples (Morris et al., 2011). Owing to considerable ground-based emissions of GNB and its ability to act as cloud condensation nuclei (CCN), these bacteria, which are plant pathogens, can influence the regional as well as global climate through cloud–aerosol interactions (Morris et al., 2011, and references therein). In particular, GNB contains β-hydroxy FAs (C₁₀–C₁₈) in lipopolysaccharides (LPS) as constituents of the outer cell membrane (Westphal, 1975). Moreover, the environmental toxic effects of GNB are, in part, due to the presence of β-hydroxy FAs present in LPS (endotoxin) (Larsson, 1994; Saraf et al., 1997; Spaan et al., 2008).

Apart from β-hydroxy FAs, other positional isomers such as α-, ω- and (ω-1)-hydroxy FAs have also been documented in various environmental archives viz. aerosols (Kawamura, 1995; Tyagi et al., 2015) and sediments (Kawamura, 1995; Wakeham et al., 2003; Zhang et al., 2014). Short chain α-hydroxy FAs (C₁₂–C₁₈) are the constituent biopolymers of fungi (Zelles, 1997), soil bacteria (Steinberger et al., 1999;...
Zelles and Bai, 1994) and protozoa (Ratledge and Wilkinson, 1988). In contrast, long chain α-hydroxy FAs (C_{16−C_{26}}) are abundant in plants, microalgae and cyanobacteria (Matsumoto and Nagashima, 1984). Likewise, ω- and (ω−1)-hydroxy FAs are highly cross-linked constituents of the cell walls of algae (Blokker et al., 1999) and plant seeds, suberin and cutin in terrestrial higher plants (Molina et al., 2006). In addition, ω- and (ω−1)-hydroxy FAs are the intermediates in the oxidation of monocarboxylic acids to dicarboxylic acids in sediments and marine aerosols (Kawamura, 1995; Kawamura and Gagosian, 1990). Furthermore, specificity of hydroxylation in FAs depends on the types of microorganisms involved (Wakeham, 1999).

These tracer compounds in snow samples may be important to better understand the contribution of plant and pathogenic bacteria to regional vs. long-range atmospheric transport (Hines et al., 2003; Lee et al., 2004, 2007; Tyagi et al., 2015) as their presence in the atmosphere can affect the CCN and ice nuclei activity (Morris et al., 2011). To the best of our knowledge, our study is the first to report α-, β- and ω-hydroxy FAs in snow samples. Snow efficiently scavenges airborne particles including soil microbes and higher plant metabolites in the free boundary layer of troposphere. Since hydroxy FAs from GNB and plants are inert in nature, they do not undergo chemical modification during snow accumulation. Therefore, hydroxy FAs in fresh snow can be used as a tracer to assess the sources and transport pathways of microorganisms and plant metabolites.

In this study, we determined hydroxy FAs in fresh snow samples collected from Sapporo, Japan, to evaluate the qualitative contribution from GNB and higher plant metabolites. Our results support the hypothesis that these hydroxy FAs are important tracers to better understand the contribution of microorganisms to the organic matter in snow. More importantly, we also discuss the possible transformations of these chemical markers during long-range atmospheric transport.

2 Experimental methods

2.1 Site description and sample collection

Sapporo (43.07°N, 141.36°E) is the capital of Hokkaido, whose population is 1.9 million (June 2013). Sapporo receives cold and dry air masses with heavy snowfall during the Asian winter monsoon. The average temperature of Sapporo in winter goes up to ~2°C (Yamamoto et al., 2011). Snow cover over the ground and fallen leaves of deciduous plants suppress the suspension of soil particles during winter, whereas the emissions of plant biomarkers from local vegetation are minimal. During the winter season, the Asian monsoon affects the regional climate, air quality and human health in Japan, delivering anthropogenic aerosols and dust from China and Siberia (Yamamoto et al., 2011). Several studies have examined the chemical and isotopic composition of ambient aerosols in various types of air masses in Sapporo (Aggarwal and Kawamura, 2008; Pavuluri et al., 2013; Yamamoto et al., 2011) to better understand the impacts of anthropogenic and biogenic contributions from Siberia, northern China and the surrounding oceans. However, no study that focuses on the transport of microorganisms using organic markers is available from Sapporo.

In this study, eleven fresh snow samples were collected from the rooftop of the Institute of Low Temperature Science (ILTS) building, Hokkaido University in Sapporo, during intensive snowfall periods (January–March) in 2010 and 2011. The detailed description about snow collection and analytical protocol of lipid fraction analyses is similar to that described in Yamamoto et al. (2011). To avoid the contribution of any possible impurities from the dry deposition of aerosols, 1–2 cm of surface snow cover were removed prior to sample collection. Thereafter, snow samples were collected in a cleaned glass jar (8 L) by using a stainless steel shovel. In each glass jar, mercuric chloride (HgCl₂) was added before sampling to prevent microbial activity. Soon after the collection, glass jars were tightened with a Teflon-lined screw cap and stored at −20°C until analysis.

2.2 Identification and quantification of hydroxy FAs

The analytical protocol used for assessing the atmospheric abundances of hydroxy FAs is described in Yamamoto et al. (2011). In brief, melted snow samples (0.5–1 L) were saponified with 1.0 M KOH in methanol at 80°C for 2 h. After saponification, the neutral fraction was separated and the remaining solution was acidified with 6 M HCl to form free carboxylic acids. Furthermore, these acids were derivatized with BF₃-methanol to form their methyl esters. The hydroxy acid methyl esters were isolated on a silica gel column by eluting with methylene chloride/methanol (95:5). The hydroxy FA methyl esters were then, derivatized to their trimethylsilyl (TMS) ethers with N,O-bis(trimethylsilyl) trifluoroacetamide (BSTFA) (SUPELCO™ Analytical) at 70°C for 1 h. After the reaction, 50μL of n-hexane solution containing 1.43 ng μL⁻¹ of internal standard (C₁₃ n-alkane/tridecane, Wako) were added to dilute the derivatives prior to GC/MS injection (Hewlett-Packard Model 6890 GC coupled to the Hewlett-Packard Model 5973 mass-selective detector, MSD). The GC was installed with a split/splitless injector and DB-5MS fused silica capillary column.

For the quantification of hydroxy FAs, the GC oven temperature was programmed from 50°C (2 min) to 305°C (15 min) at 5°C min⁻¹. Data were acquired and processed with the Chemstation software. Structural identification and comparison of retention time of hydroxy FAs were performed using authentic TMS derivatives of n-C_{12} and n-C_{16} α-hydroxy FAs, n-C_{12}, n-C_{14}, n-C_{15}, and n-C_{16} β-hydroxy FAs and n-C_{16}, n-C_{20} and n-C_{22} ω-hydroxy FAs. The recoveries of authentic fatty acid standards were better than
92 ± 4% with analytical error (average 4.1%) for acidic compounds (Yamamoto et al., 2011). Laboratory blanks showed no contamination of any target compounds. The results of n-alkanes, n-alkanols and n-alkanoic acids (terrestrial biomarkers) in snow samples are reported in Yamamoto et al. (2011), which revealed a long-range atmospheric transport of terrestrial organic materials from northeastern Asia to northern Japan by the Asian winter monsoon.

2.3 Estimation of endotoxin levels and mass loading of GNB

Since the endotoxins from GNB contain β-hydroxy FAs from C_{10} to C_{18}, previous studies attempted to quantify atmospheric abundances of endotoxins using the concentrations of ambient hydroxy FAs measured (Lee et al., 2004; Rietschel et al., 1984; Wilkinson, 1988). According to these studies, concentrations of endotoxins in snow samples can be estimated from the mathematical expression as below.

Endotoxins (LPS, ng kg\(^{-1}\) of meltwater) = 
\[
\left( \Sigma \beta\text{-hydroxy FAs from } C_{10} \text{ to } C_{18}; \text{nmol kg}^{-1} \right) \times 8000 
\] 
\[/4
\]

In the above formula, the average molecular weight of endotoxin corresponds to 8000 as reported by Mielniczuk et al. (1993). β-Hydroxy FAs in the mathematical expression are the total (LPS-bound + free) hydroxy FAs for the carbon numbers from C_{10} to C_{18}. We also estimated the mass loading of airborne GNB using the approach initially suggested by Balkwill et al. (1988) and later on by Lee et al. (2004), in which they used the chemical marker to bacterial mass conversion factor of 15 nmol of β-hydroxy FAs (C_{10}–C_{18}) per mg dry cell weight. Therefore, we have converted the sum of mass concentrations of β-hydroxy FAs from C_{10} to C_{18} (in nmol kg\(^{-1}\)) into the equivalent dry cell weight of GNB (i.e., in mg kg\(^{-1}\) of meltwater) by normalizing with 15.

3 Results and discussion

3.1 Air mass backward-trajectory analysis

The air mass back trajectories (AMBTs) provide a means to qualitatively assess the source regions of airborne pollutants over a receptor site. For this study, we have computed 7-day isentropic AMBTs using the Hybrid Single Particle Lagrangian Integrated Trajectory (HYSPLIT) model (Draxler and Rolph, 2013, and references therein). The meteorological parameters (GDAS data sets) from the NOAA Air Resources Laboratory were used as an input for the HYSPLIT model. Figure 1 shows the AMBT cluster at an arrival height of 500 m a.g.l. (above ground level) for the sampling days in (a) winter 2010 and (b) winter 2011.

Figure 1. Air mass back-trajectory cluster at an arrival height of 500 m a.g.l. (above ground level) for the sampling days in (a) winter 2010 and (b) winter 2011.

3.2 Concentrations of hydroxy fatty acids

Homologous series of α-, β- and ω-hydroxy FAs were detected in fresh snow samples collected from Sapporo. Their mass concentrations are summarized in Tables 1 and 2 for the winters of 2010 and 2011, respectively. Based on 2-year seasonal data on hydroxy FAs, we found that concentrations of α-hydroxy FAs are significantly higher than β- and ω-hydroxy FAs. The predominance of α-hydroxy FAs can be explained by the α-oxidation pathway of FAs, which generally occurs in plants, animals and bacteria (Cranwell, 1981, and references therein) whereas β- and ω-oxidation is specific to bacteria (Lehninger, 1975). α-Hydroxy FAs, in particular high molecular weight ones, come from the epicuticular waxes of higher plants as well as from algae. However, we also found higher abundance of α-hydroxy FAs in the biomass burning aerosols collected over Mt. Tai, China (Tyagi et al., 2015), possibly due to photochemical oxidation of higher molecular weight fatty acids. Such a possibility of in situ formation of α-hydroxy FAs has also been
reported in the hydrolysis products of leaf waxes and wood, and in microalgae and sea grasses (Feng et al., 2015). Furthermore, microbial oxidation could also be a possible source of \( \alpha \)-hydroxy FAs (Eglington et al., 1968) in the snow samples studied. Hence, we suggest that \( \alpha \)-hydroxy FAs cannot be employed as the tracers of plant waxes only, as they can come from microbial/photocatalytic oxidation of higher molecular weight fatty acids during long-range atmospheric transport.

A characteristic feature of our data is the predominance of \( C_{16} \) hydroxy FAs in all the types of hydroxy FAs measured. However, significant shifts were observed in the carbon numbers of the second-most abundant \( \beta \)-hydroxy FAs (mostly C-number > 16) and \( \omega \)-hydroxy FAs (i.e., C-number < 16; see Tables 1 and 2). A likely explanation for this observation is that \( \beta \)-hydroxy FAs above \( C_{16} \) were formed by \( \beta \)-oxidation of long chain FAs, which are more common in microorganisms as discussed previously. In contrast, \( \omega \)-hydroxy FAs below \( C_{16} \) are present in plants and microbes (Cardoso and Eglington, 1983), in which \( \omega \)-oxidation of fatty acids is the secondary choice for microbial oxidation.

### 3.3 Molecular distributions

Figure 2 presents molecular distributions of \( \alpha \)- (\( C_{9} \) to \( C_{30} \)), \( \beta \)- and \( \omega \)-hydroxy FAs (\( C_{9} \) to \( C_{28} \)) in snow samples from Sapporo during the winter of 2010 and 2011. Even carbon number predominance is noteworthy for \( \alpha \)-, \( \beta \)- and \( \omega \)-hydroxy FAs. \( \alpha \)-Hydroxy FAs show molecular distributions with the order \( C_{16} > C_{24} > C_{22} \) in both years (see Fig. 2a). Likewise, \( \beta \)-hydroxy FAs show the predominance of \( C_{16} \) followed by \( C_{18} \) or \( C_{20} \) and then by \( C_{14} \) in both winters. However, we found the predominance of \( C_{20} \) \( \beta \)-hydroxy FAs over \( C_{16} \) in one snow sample during 2010. Similarly, \( \omega \)-hydroxy FAs showed dominance of \( C_{16} \) followed by the others as \( C_{14} > C_{12} \sim C_{22} \sim C_{24} \) during snowfall in both the years.

Table S1 in the Supplement describes the statistically significant differences in the ratios of even to odd carbon numbers for \( \alpha \)-, \( \beta \)-, and \( \omega \)-hydroxy FAs in snow samples based on a two-tailed unpaired \( t \) test. No significant differences were observed between 2010 and 2011 for the ratios of even to odd carbon number \( \alpha \)-hydroxy FAs. In contrast, the difference is statistically significant between 2010 and 2011 for \( \beta \)- and \( \omega \)-hydroxy FAs. In fact, the difference is much larger for \( \omega \)-hydroxy FAs than that for \( \beta \) isomers. In the 2010
winter, AMBTs show atmospheric transport from the continents at 500, 1000 and 1500 m above ground; however, at the same heights in the 2011 winter, the air masses came from the oceans during one sample collection. Higher plants in the continents contribute to higher abundances of hydroxy FAs than the oceans, and thus explain higher abundances of \( \beta \)- and \( \omega \)-hydroxy FAs in 2010 than in 2011. On average, even carbon numbered \( \alpha \)-, \( \beta \)- and \( \omega \)-hydroxy FAs in their total mass concentrations account for \( \sim 0.67 \) ng kg\(^{-1}\), suggesting as a significant source of \( C_{16} \) to \( C_{22} \) \( \alpha \)-, \( \beta \)- and \( \omega \)-hydroxy FAs (Cardoso and Eglinton, 1983). In a similar way, it has been proposed that hydroxy FAs \( (C_{20}-C_{30}) \) are principally derived from terrestrial higher plants (Kawamura and Ishiwatari, 1984). Therefore, \( \alpha \)-, \( \beta \)- and \( \omega \)-hydroxy FAs \( (C_{16}-C_{22}) \) in snow samples can be related to their sources from terrestrial higher plants through long-range atmospheric transport.

Previous studies documented ubiquitous occurrence of these hydroxy FAs in soil microbes such as yeast and fungi (Van Dyk et al., 1994, and references therein) and in the LPS of GNB (Lee et al., 2007). In this regard, prior studies focusing on \( \beta \)-hydroxy FAs with the predominance of \( C_{16} \) and \( C_{18} \) suggested the contributions from yeast and fungi (Stodola, 1967; Van Dyk et al., 1994, and references therein). Molecular distributions of \( \beta \)-hydroxy FAs show a predominance of \( C_{16} \) followed by \( C_{18} \) or \( C_{20} \) (see Fig. 2b), suggesting that they have been derived from soil microbes. Likewise, FAs \( < C_{20} \) are derived from marine phytoplankton (Kawa-

### Table 2. Mass concentrations (in ng kg\(^{-1}\)) of \( \alpha \)-, \( \beta \)- and \( \omega \)-hydroxy fatty acids (FAs) measured in snow samples \((N = 6)\) collected from Sapporo during the winter of 2011.

<table>
<thead>
<tr>
<th>C-number</th>
<th>( \alpha )-Hydroxy FAs</th>
<th>( \beta )-Hydroxy FAs</th>
<th>( \omega )-Hydroxy FAs</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Range</td>
<td>Mean ± S.E. Median</td>
<td>Range</td>
</tr>
<tr>
<td>C_{9}</td>
<td>b.d.–27.2</td>
<td>14.2 ± 5.7</td>
<td>1–8.5</td>
</tr>
<tr>
<td>C_{10}</td>
<td>b.d.–65.4</td>
<td>30.9 ± 11.2</td>
<td>1.7–12.7</td>
</tr>
<tr>
<td>C_{11}</td>
<td>19.8–66.6</td>
<td>34.2 ± 8.5</td>
<td>1.7–13.3</td>
</tr>
<tr>
<td>C_{12}</td>
<td>20.7–60.4</td>
<td>36.5 ± 6.6</td>
<td>1.3–15.3</td>
</tr>
<tr>
<td>C_{13}</td>
<td>b.d.–49.2</td>
<td>21.5 ± 8.2</td>
<td>4.5–15.8</td>
</tr>
<tr>
<td>C_{14}</td>
<td>7.5–55.3</td>
<td>28.6 ± 7.7</td>
<td>4.5–25.5</td>
</tr>
<tr>
<td>C_{15}</td>
<td>b.d.–77.6</td>
<td>29.2 ± 13.1</td>
<td>1.9–11.1</td>
</tr>
<tr>
<td>C_{16}</td>
<td>14.3–186</td>
<td>94.0 ± 29.3</td>
<td>2.8–55.8</td>
</tr>
<tr>
<td>C_{17}</td>
<td>2.8–29.3</td>
<td>15.3 ± 4.3</td>
<td>1.6–12.2</td>
</tr>
<tr>
<td>C_{18}</td>
<td>8.0–55.8</td>
<td>31.3 ± 8.2</td>
<td>0.6–31.4</td>
</tr>
<tr>
<td>C_{19}</td>
<td>b.d.–22.4</td>
<td>6.2 ± 4.4</td>
<td>1.9–10.9</td>
</tr>
<tr>
<td>C_{20}</td>
<td>11.5–97.9</td>
<td>53.5 ± 18.6</td>
<td>1.2–43.4</td>
</tr>
<tr>
<td>C_{21}</td>
<td>b.d.–95.2</td>
<td>29.1 ± 17.2</td>
<td>1.0–16.6</td>
</tr>
<tr>
<td>C_{22}</td>
<td>13.4–109</td>
<td>60.8 ± 19.9</td>
<td>1.6–27.2</td>
</tr>
<tr>
<td>C_{23}</td>
<td>8.1–58.1</td>
<td>32.2 ± 10.1</td>
<td>5.7–11.6</td>
</tr>
<tr>
<td>C_{24}</td>
<td>12.3–92.2</td>
<td>74.9 ± 34</td>
<td>19.1–24.3</td>
</tr>
<tr>
<td>C_{25}</td>
<td>2.6–51.3</td>
<td>18.4 ± 8.9</td>
<td>3.3–11.1</td>
</tr>
<tr>
<td>C_{26}</td>
<td>2.6–52.0</td>
<td>24.2 ± 9</td>
<td>b.d.–15.9</td>
</tr>
<tr>
<td>C_{27}</td>
<td>b.d.–5.6</td>
<td>2 ± 1.3</td>
<td>b.d.–9.2</td>
</tr>
<tr>
<td>C_{28}</td>
<td>b.d.–4.8</td>
<td>1.4 ± 0.9</td>
<td>b.d.–10.6</td>
</tr>
<tr>
<td>C_{29}</td>
<td>b.d.–3.35</td>
<td>0.7 ± 0.67</td>
<td>0</td>
</tr>
<tr>
<td>C_{30}</td>
<td>b.d.–0.60</td>
<td>0.12 ± 0.12</td>
<td>0</td>
</tr>
</tbody>
</table>

|          | Total                   | 619–1279               | 639 ± 187              | 651                     | 6–354                  | 179 ± 64                  | 170                     | 27–422                 | 149 ± 73               | 102                    |

Note: b.d. = below detection limit \( \leq 0.06 \) ng kg\(^{-1}\). S.E. (standard error) = \( \sigma / \sqrt{N} \), where \( \sigma \) refers to the standard deviation of total samples \((N)\).
Figure 2. Molecular distributions of (a) α-hydroxy fatty acids (FAs) (C₉–C₃₀), (b) β-hydroxy FAs (C₀–C₂₈) and (c) ω-hydroxy FAs (C₀–C₂₈) in the snow samples collected from Sapporo during the winter of 2010 and 2011.

Figure 3 depicts bar graphs showing the relative abundances of α-, β- and ω-hydroxy FAs in the snow samples from Sapporo during winter. We found that the proportions of two classified groups (low molecular weight C₀–C₁₉ and high molecular weight C₂₀–C₃₀ or C₂₀–C₂₈) of α-, β- and ω-hydroxy FAs are very similar between 2010 and 2011 (Fig. 3). This observation is perhaps related to their common sources/transport pathways of α-, β- and ω-hydroxy FAs over Sapporo. This inference is further supported by the AMBTs computed at arrival heights of 500, 1000 and 1500 m (see Figs. 1 and S1 in the Supplement), indicating similar air mass transport pathways from Russia and Siberia.

3.4 Endotoxin potency of GNB impact via aeolian transport

Endotoxin in GNB determines their viability and potentially causes pathological effects on mammals (Lüderitz et al., 1981; Westphal, 1975). In particular, GNB contain LPS in their outer membrane. When bacteria multiply, die and lyse, LPS are released from the surface as a potential bacterial toxin, and are therefore called an endotoxin (Westphal, 1975). In addition to intact bacterial cells, this endotoxin can trigger and cause allergies, respiratory problems and infections. Researchers have used LPS concentrations as a measure of GNB, primarily by the limulus amebocyte lysate (LAL) assay that has limited specificity (Saraf et al., 1997). The β-hydroxy FAs, markers for endotoxin/LPS, were assayed in various environmental samples such as dust (Andersson et al., 1999; Hines et al., 2000), aerosols (Lee et al., 2004, 2007; Walters et al., 1994), soils (Keinänen et al., 2003), sewage (Spaan et al., 2008) and marine dissolved organic matter (Wakeham, 1999).
As mentioned in Sect. 2.3, we have estimated the abundances of endotoxin and mass loading of GNB in fresh snow samples. This quantification is indeed crucial for assessing a likely allergic impact of endotoxin globally via long-range atmospheric transport. Here, we estimated the endotoxin concentrations in snow as varying from 424 to 1080 ng kg$^{-1}$ (ave. 789 ± 237 ng kg$^{-1}$) in 2010 and from 36 to 1100 ng kg$^{-1}$ (ave. 579 ± 435 ng kg$^{-1}$) in 2011 samples. The estimated lower limits of endotoxin in Tables 1 and 2 are calculated based on the minimum concentration of β-hydroxy FAs (C$_{10}$−C$_{18}$), which are specific to Gram-negative bacteria (GNB). β-Hydroxy FAs (C$_{10}$−C$_{18}$) are the structural constituents of lipid A, which are present in the outer cell membrane of GNB. Thus, the endotoxin concentrations in snow samples were estimated based on the abundances of β-hydroxy FAs having a carbon chain length from 10 to 18 (Sect. 2.3). Consistent with this study, Lee et al. (2004) also reported endotoxin concentrations based on β-hydroxy FAs (C$_{10}$−C$_{18}$). Although relative abundances of endotoxin during the winter of 2010 (N = 5) are higher than those of 2011 samples (N = 6), the two-tailed t test revealed no significant differences (t = 0.96; df = 9; P > 0.05) with regard to mean concentrations of the 2 years.

In this study, we estimated dry mass concentrations of GNB in snow samples to be 26.3 ± 7.9 µg kg$^{-1}$ in 2010 vs. 19.3 ± 1.4 µg kg$^{-1}$ in 2011. Lee et al. (2007) reported that airborne endotoxin is of crustal origin and thus can be transported long distances to the outflow region. Since the AMBTs reveal the impact of long-range transport from Russia and Siberia during the study period, we infer that estimated endotoxin concentrations and dry cell weight of GNB over Sapporo are derived from those source regions. Recently, Golokhvast (2014) documented the airborne biogenic particles in snow from the Russian Far East that cause allergy for the pedestrians. The airborne biogenic particles can be scavenged efficiently by both wet precipitation and snowfall. Therefore, we have looked for the literature describing the occurrence of GNB in rainwater for comparison with our study on Sapporo snow. Towards this, Gould (1999) and Lye (2002) have documented the presence of various GNB (e.g., Salmonella, Shigella, Vibrio, Legionella and Campylobacter spp.) species in rainwater. Likewise, Kawamura and Kaplan (1983) also reported the presence of β-hydroxy FAs in rainwater samples collected from Los Angeles (USA) and attributed their sources to bacterial membranes. So far, no literature has been available on endotoxin and GNB concentrations in snow samples from East Asia in order to make a comprehensive comparison with the present study.

Overall, the presence of endotoxin and GNB in snow affirms that biogenic particles of soil microbes and their potential health impact should not be overlooked. Routine and long-term measurements of airborne chemical markers (hydroxy FAs in this study) could aid the monitoring of the microbial content in long-range transported air masses. Further studies are required to examine their distributions in the atmospheric environment and health effects on human beings in the regional and global perspectives during long-range atmospheric transport.

4 Conclusions

Although low temperature is considered to be a limiting factor for bacterial activity in air/snow, some studies have shown that bacteria can be metabolically active even at subzero temperatures (Polymenakou, 2012, and references therein). Figure 4 summarizes the whole idea, which was addressed in this study. We conclude that fresh snow in Japan acts as a filter for bacterial activity in air/snow, some studies have shown that bacteria can be metabolically active even at subzero temperatures (Polymenakou, 2012, and references therein). Figure 4 summarizes the whole idea, which was addressed in this study. We conclude that fresh snow in Japan acts as a filter.
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P. Tyagi conducted the experiments. P. Tyagi prepared the manuscript with contributions from K. Kawamura.

Author contributions. S. Yamamoto extracted the samples and conducted the experiments. P. Tyagi prepared the manuscript with contribution from K. Kawamura.

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