



## Macroalgae contribute to nested mosaics of pH variability in a subarctic fjord

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**Abstract.** The Arctic Ocean is considered the most vulnerable ecosystem to ocean acidification, and large-scale assessments of pH and the saturation state for aragonite ( $\Omega_{\text{arag}}$ ) have led to the notion that the Arctic Ocean is already close to a corrosive state. In high-latitude coastal waters the regulation of pH and  $\Omega_{\text{arag}}$  is, however, far more complex than offshore because increased biological activity and input of glacial meltwater affect pH. Effects of ocean acidification on calcifiers and non-calcifying phototrophs occupying coastal habitats cannot be derived from extrapolation of current and forecasted offshore conditions, but they require an understanding of the regimes of pH and  $\Omega_{\text{arag}}$  in their coastal habitats. To increase knowledge of the natural variability in pH in the Arctic coastal zone and specifically to test the influence of benthic vegetated habitats, we quantified pH variability in a Greenland fjord in a nested-scale approach. A sensor array logging pH,  $\text{O}_2$ , PAR, temperature and salinity was applied on spatial scales ranging from kilometre scale across the horizontal extension of the fjord; to 100 m scale vertically in the fjord, 10–100 m scale between subtidal habitats with and without kelp forests and between vegetated tidal pools and adjacent vegetated shores; and to centimetre to

metre scale within kelp forests and millimetre scale across diffusive boundary layers of macrophyte tissue. In addition, we assessed the temporal variability in pH on diurnal and seasonal scales. Based on pH measurements combined with point samples of total alkalinity, dissolved inorganic carbon and relationships to salinity, we also estimated variability in  $\Omega_{\text{arag}}$ . Results show variability in pH and  $\Omega_{\text{arag}}$  of up to 0.2–0.3 units at several scales, i.e. along the horizontal and vertical extension of the fjord, between seasons and on a diel basis in benthic habitats and within 1 m<sup>3</sup> of kelp forest. Vegetated intertidal pools exhibited extreme diel pH variability of > 1.5 units and macrophyte diffusive boundary layers a pH range of up to 0.8 units. Overall, pelagic and benthic metabolism was an important driver of pH and  $\Omega_{\text{arag}}$  producing mosaics of variability from low levels in the dark to peak levels at high irradiance generally appearing favourable for calcification. We suggest that productive coastal environments may form niches of high pH in a future acidified Arctic Ocean.

## 1 Introduction

The Arctic Ocean is considered to be the most vulnerable ecosystem to ocean acidification (OA) due to the combined effects of low temperature, which increases the solubility of CO<sub>2</sub> and, in places, dilution of the buffering capacity of seawater by freshwater inputs (Fabry et al., 2009; AMAP, 2013). Indeed, large-scale assessments of pH in combination with saturation states for aragonite ( $\Omega_{\text{arag}} < 1$ ) have led to the notion that the Arctic Ocean is already close to a corrosive state (Fabry et al., 2009). However, whereas this has been documented for offshore waters, the Arctic contains a massive coastline where the regulation of pH and  $\Omega_{\text{arag}}$  is far more complex than that offshore (Hofmann et al., 2011; Duarte et al., 2013). In coastal waters, the role of air–sea CO<sub>2</sub> exchange in regulating pH operates along with watershed effects driven by the discharge of freshwater and the effects of metabolically intense communities on pH (Duarte et al., 2013). The Greenland Ice Sheet is melting at a rate that has more than doubled in the last decade (Helm et al., 2014) and Greenland fjords are hence potentially among the most susceptible to the effects of freshening and acidification.

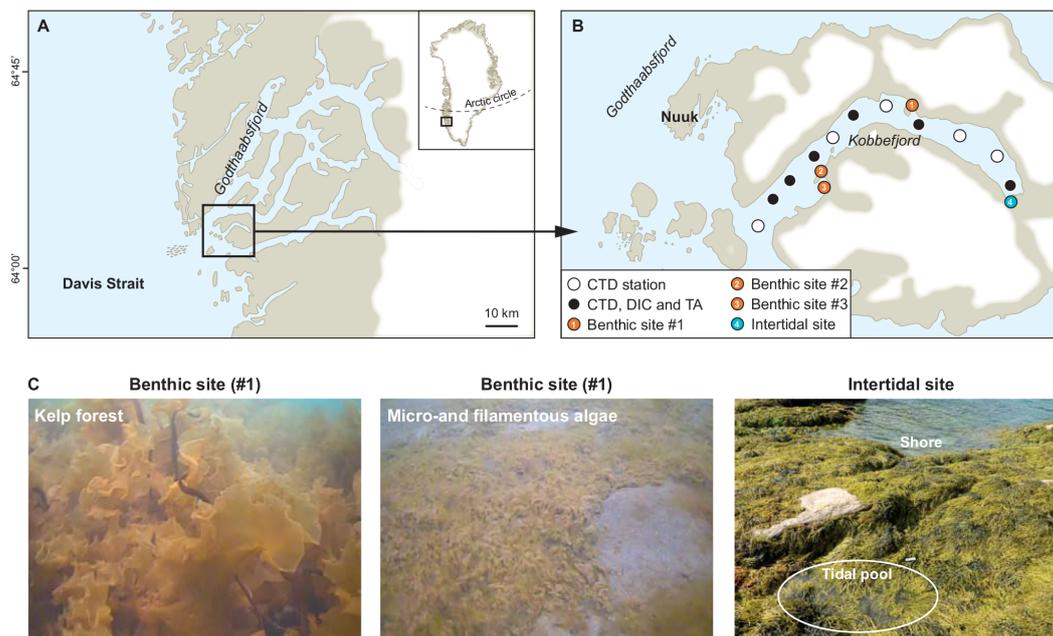
As most calcifiers occupy coastal habitats, the assessment of risks of Arctic acidification to these vulnerable species cannot be derived from extrapolation of the current and forecasted offshore conditions alone, requiring instead an understanding of the regimes of pH and  $\Omega_{\text{arag}}$  in the coastal habitats they occupy, and the same is true regarding potential effects of ocean acidification on coastal phototrophs (calcifying or non-calcifying) (Mercado and Gordillo, 2011). Such information is currently largely lacking for the Arctic in general and for Greenland in particular, which calls for efforts to understand variability in pH in the coastal zone informing on the factors controlling pH and ultimately determining the sensitivity of the coastal Arctic Ocean ecosystem to ocean acidification.

Greenland has a vast and highly indented coastline, extending approximately 44 000 km and representing ca. 12 % of the world's coastline (Krause-Jensen and Duarte, 2014). This coastline forms a complex network of fjords and open coasts that contains multiple features contributing to heterogeneity, such as continental ice and freshwater discharge at the headwaters, variable slopes and substrates, differential water residence time conducive to widely distinct temperature regimes within neighbouring areas (Olesen et al., 2015), and tides that generate intertidal habitats and force flow patterns. In addition, Greenland fjords often support highly productive kelp forests (Krause-Jensen et al., 2012) and intertidal seaweed communities (Høgslund et al., 2014), which have been suggested to have the capacity to affect pH and  $\Omega_{\text{arag}}$  locally (Krause-Jensen and Duarte, 2014). Such effects have been demonstrated for Antarctic and temperate kelp/macroalgal ecosystems (Middelboe and Hansen 2007; Delille et al., 2009; Cornwall et al., 2013a) as well as for subtropical and tropical seagrass meadows (e.g. Hofmann et

al., 2011; Hendriks et al., 2014). Calcifiers such as bivalves, brittle stars and sea urchins, which are potentially vulnerable to OA, are ecologically important as they contribute significantly to carbon cycling in both the subarctic and Arctic Greenland, where their distribution ranges from the intertidal zone to > 300 m depth (Sejr et al., 2002; Blicher et al., 2007, 2009, 2013; Blicher and Sejr, 2011). Phototrophs such as kelps, while being able to affect the pH regime, may also respond to OA, which has been shown to stimulate their growth (Olischläger et al., 2012) and affect the competition between kelps and understory red algae (Connell and Russell 2010).

Although the variability in pH and  $\Omega_{\text{arag}}$  in Greenland fjords has not been reported, available oceanography and environmental surveys suggest that this may be substantial. For instance, in Young Sound, Sejr et al. (2011) found that the extent of sea-ice cover and inputs of glacial melt water affect seawater  $p\text{CO}_2$  levels and sea–air exchange at spatial, seasonal and interannual scales. Seasonal dynamics of autotrophic and heterotrophic plankton metabolism have also been found to markedly affect  $p\text{CO}_2$  levels in Kobbefjord, a subarctic fjord in SW Greenland (Sejr et al., 2014). However, information on scales of variability in pH and  $\Omega_{\text{arag}}$  in Greenland fjords is still lacking, precluding the assessment of their current and future vulnerability to ocean acidification.

Here we quantify pH variability in Kobbefjord, SW Greenland. This subarctic fjord supports dense and productive subtidal kelp forests, intertidal macroalgal habitats and high abundance of bivalves and sea urchins with important roles in the ecosystem (Blicher et al., 2009; Krause-Jensen et al., 2012). We hypothesize that Kobbefjord contains a mosaic of pH environments nested across a range of scales of variability and that primary production in general, and by macroalgae in particular, may be an important driver of pH variability relevant for benthic calcifiers. We first assess seasonal and spatial variability in the open-water pH at the kilometre scale along the horizontal extension and at 100 m scale vertically in the fjord. We then examine diel variability in pH within subtidal benthic habitats colonized by kelp forest or microalgae/scattered filamentous algae as well as in vegetated tidal pools and adjacent vegetated intertidal shores, with the distance between parallel deployments at the 10–100 m scale. We further explore the pH variability three-dimensionally at centimetre to metre scale within the kelp forest ecosystem and at millimetre scale across the diffusive boundary layer (i.e. the layer in which molecular diffusion is the dominant transport mechanism for dissolved material; see e.g. de Beer and Larkum, 2001) of key macrophyte species. Whereas our assessment focuses on pH, we also discuss the associated variability in  $\Omega_{\text{arag}}$ .



**Figure 1.** (a) Location of Kobbefjord, Nuuk. (a) Location of sampling sites in Kobbefjord: fjord-scale sites (CTD,  $C_T$ ,  $A_T$ : filled circles; CTD: open circles), vegetated subtidal sites (open circles #1–3), and intertidal sites (open circles #4). (c) Photopanel of benthic habitats: a typical kelp forest habitat (dominated by *Saccharina longicruris*) and habitat colonized by microalgae/scattered filamentous algae (example from site #1, representative of sites #1–3 in map) and a vegetated intertidal pool and the adjacent vegetated shore dominated by *Ascophyllum nodosum* and *Fucus* spp. (site #4 in map).

## 2 Methods

### 2.1 Study area

Kobbefjord is located in the extensive Godthåbsfjord system in southwest Greenland (Fig. 1a). The fjord is 17 km long and 0.8–2 km wide and has a maximum depth of 150 m. It is subjected to marked exchange of coastal water driven by a tidal range of 1–4.5 m (Richter et al., 2011) and receives freshwater mainly from a river in the innermost part of the fjord, leading to a salinity gradient in the surface water. Sea ice usually covers the inner part of the fjord from December to early May, but the outer part of the fjord is permanently ice-free. Light attenuation in the water column has been reported to range from  $0.083\text{ m}^{-1}$  in February to  $0.197\text{ m}^{-1}$  in May to  $0.135\text{ m}^{-1}$  in September (Sejr et al., 2014). Whereas the phytoplankton community is the main primary producer in the central parts of the fjord (Sejr et al., 2014), subtidal macroalgae, dominated by *Saccharina longicruris* and *Agarum clathratum*, form productive benthic habitats along the shores to water depths of ca. 40 m (Krause-Jensen et al., 2012) interspaced with communities of benthic microalgae (Glud et al., 2010; Attard et al., 2014) as well as with scattered eelgrass (*Zostera marina*) meadows at 1–3 m depth (Olesen et al., 2015). Communities of intertidal macroalgae, dominated by *Fucus* spp. and *Ascophyllum nodosum*, are prominent in the intertidal zone, where they form

an important habitat for blue mussels, for example (Blicher et al., 2013).

Three field campaigns targeting seasonal- and fjord-scale variability in pH in the pelagic zone were conducted in the spring (19 April), midsummer (18 July) and late summer (3 September) of 2013 (Fig. 1b). The late summer survey was associated with an intensive campaign (27 August–6 September 2013) exploring pH variability in shallow subtidal kelp habitats and neighbouring habitats colonized by benthic microalgae and scattered filamentous algae (Fig. 1c). A final late summer campaign (22–30 August 2014) addressed pH variability in vegetated tidal pools and surface waters of adjacent vegetated shores (Fig. 1c). All pH data from fjord-scale to microscale are reported on the total pH scale.

### 2.2 Fjord- and seasonal-scale pH variation

To determine the large-scale spatial and seasonal variation in physical and chemical parameters in the water column of Kobbefjord, vertical profiles were performed at 11 stations located along a longitudinal gradient following the main central axis of the fjord on 19 April, 18 July, and 3 September 2013 (Fig. 1b). We used a Sea-Bird CTD (SBE 19plus) equipped with sensors for temperature, conductivity, fluorescence (Seapoint chlorophyll fluorometer), oxygen (SBE 43, Sea-Bird) and pH (SBE 18, Sea-Bird). Alongside CTD profiles, water samples were collected using a 5 L Niskin bottle at 1, 5, 10, 20, 30, and 40 m depth. Water was collected

for dissolved oxygen measurement using Winkler titration (Parsons et al., 1984), which was used to calibrate the CTD oxygen optode. The pH sensor was calibrated using NIST buffers and a seawater Tris buffer prepared according to Dickson (2007). Unfiltered water was transferred to 150 mL borosilicate glass bottles for pH analysis. The samples were poisoned with a saturated mercuric chloride solution, cooled and stored in darkness until arrival. Back in the lab, pH was measured potentiometrically using a glass reference electrode (Orion, Ross Ultra pH/ATC Triode) calibrated with NIST buffers and a seawater Tris buffer prepared according to Dickson (2007). The measurements were used to correct the offset of the SBE 18 pH measurements.

For estimation of the saturation state of aragonite ( $\Omega_{\text{arag}}$ ), samples for analyses of dissolved inorganic carbon ( $C_T$ ) and total alkalinity ( $A_T$ ) were collected at five stations on one occasion (3 September 2013). Triplicate 12 mL samples were collected at 5, 10, 20, 30, and 40 m depth and near the bottom. Samples were carefully siphoned through Tygon tubing from Niskin bottles to 12 mL septum-capped glass vial (Exetainers), allowing the water to overflow for two volume changes. The samples were poisoned with 100  $\mu\text{L}$  5 %  $\text{HgCl}_2$  to avoid biological alteration.  $C_T$  was analysed with a  $C_T$  analyser (AS-C3, Apollo SciTech Inc). The accuracy of the analysis was 2.4  $\mu\text{mol kg}^{-1}$  (average numerical deviation from the reference material value) and the precision was 1.4  $\mu\text{mol kg}^{-1}$  (average standard deviation of triplicate samples).  $A_T$  was analysed on an alkalinity titrator (AS-ALK2 from Apollo SciTech), with verification against the same certified reference material used for pH measurements or a Metrohm Titrando 808 by open-cell titration (Dickson et al., 2007) using Batch 136 supplied by the Andrew Dickson lab at UC San Diego for verification. Average analysis accuracy was 2.9  $\mu\text{mol kg}^{-1}$  (average numerical deviation from the reference material value). Relationships between the point samples of  $A_T$  and salinity ( $S$ ) were used to verify the published relationship for the Godthåbsfjord system ( $\text{TA} = 159 + 63 S$ ; Meire et al., 2015), which was subsequently applied for estimation of  $A_T$  for the full September data set.  $\Omega_{\text{arag}}$  and  $p\text{CO}_2$  were calculated from  $A_T$  and pH using the  $\text{CO}_2\text{SYS}$  Excel program version 2.1 (Pierrot et al., 2006) with the  $K_1$  and  $K_2$  constants from Mehrbach et al. (1973), as modified by Dickson and Millero (1987).

### 2.3 Small-scale and diurnal-scale pH variation

To measure small-scale and diurnal-scale variation in pH and physico-chemical variables in kelp forests and adjacent subtidal habitats colonized by microalgae and scattered filamentous algae we constructed metal frames measuring approximately 0.90 m  $\times$  0.90 m  $\times$  1.10 m. Each frame was equipped with instruments that allowed continuous measurements of temperature, salinity, water level, oxygen concentration, photosynthetically active radiation (PAR) and pH at ca. 50 cm above the seafloor (Fig. A1). Measurements were made ev-

ery 10 min or less. We selected three dense (close to 100 % cover) kelp beds located in shallow water (average depth 2–5 m) in different sites of the fjord. All kelp beds were dominated by *S. longicuris* with co-occurrence of *A. clathratum* and were surrounded by habitats colonized by microalgae and varying amounts of scattered filamentous algae. We conducted parallel deployments of frames with loggers in kelp beds vs. surrounding non-kelp habitats in each of the three sites, with each deployment lasting about 48 h. The typical distance between kelp and non-kelp habitats at each site was approximately 100 m. Conductivity, temperature and water level were measured with a Hydrolab DS5X and a MicroCAT (SBE37, Sea-Bird). Oxygen concentration was measured using miniDOT oxygen loggers (Precision Measurement Engineering) and a Hydrolab DS5X. PAR was measured using Odyssey PAR loggers from Dataflow Systems Pty Limited. pH was measured using Hydrolab DS5X and SeaFET pH loggers from Satlantic. Hydrolab DS5X pH sensors were calibrated with a routine two-point calibration using NIST buffers of  $\text{pH}_{\text{NBS}}$  7.0 and 10.0. Before and after each deployment all instruments were placed in a 50 L tank with seawater to intercalibrate sensors. All pH loggers were offset to the same newly calibrated high-precision SeaFET pH sensor, calibrated at the Satlantic facility ([www.satlantic.com](http://www.satlantic.com)) on the total scale using single-point calibration. Oxygen sensors were calibrated to  $\text{O}_2$  concentrations of the tank as determined from Winkler titrations.

To monitor three-dimensional pH variations on a metre scale within the kelp canopy, we deployed a custom-built multi-sensor array, consisting of an autonomous data logger (dataTaker DT85) in a watertight housing (custom-built by Albatros Marine Technologies S.L.) with 16 pre-amplified pH electrodes (Omega, PHE-1304-NB). The pH sensors were attached to the submersible logger by 5 m long cables to allow for adjusting their position as needed (Fig. A1 in Appendix). The sensors were configured in situ in a three-dimensional array on the metal frame occupying a volume of approximately 1 m<sup>3</sup>, with four sensors at 0.1 m from the bottom, four sensors at 0.2 m, four sensors just underneath the canopy and four above the canopy, which typically extended about 0.75 m above the seafloor. All pH sensors were calibrated with a three-point calibration using NIST buffers of  $\text{pH}_{\text{NBS}}$  4.0, 7.0 and 10.0, allowing at least 5 min between every reading for the sensors to stabilize. All pH loggers were offset to the same newly calibrated high-precision SeaFET pH sensor as mentioned above. On several occasions triplicate samples for determination of  $C_T$  and  $A_T$  were collected and analysed as described above to allow calculation of carbonate chemistry and  $\Omega_{\text{arag}}$ .

pH variation in vegetated tidal pools dominated by *Ascophyllum nodosum* and adjacent intertidal habitats on the shore also dominated by *A. nodosum* and *Fucus* spp. was quantified over a diurnal cycle through sampling at low tide just after pool formation and prior to pool inundation during day and night. pH and  $\Omega_{\text{arag}}$  were calculated from  $C_T$  and  $A_T$

samples collected and analysed as described above and computed using the CO2SYS program (Pierrot et al., 2006) with in situ information on temperature and salinity. Salinity was analysed from water samples based on measurements of conductivity (Orion 3-Star conductivity benchtop), while oxygen concentration and water temperature were determined using a portable meter (Hack, HQ40d).

## 2.4 Microscale pH variation

pH variations at a millimetre scale were measured in the laboratory on six different species of macrophytes (the intertidal brown macroalgae *Ascophyllum nodosum* and *Fucus vesiculosus*, the kelps *Saccharina longicruris* and *Agarum clathratum*, the green alga *Ulva lactuca*, and the seagrass *Zostera marina*) occurring in Kobbefjord and collected either there or, for logistic reasons, in another branch of the Godthåbsfjord system. From each species, a piece of approximately  $5 \times 2$  cm was cut and mounted on a microscope slide in an aquarium with seawater before measurements. The setup was mounted in an aquarium in a climate-controlled room with temperature kept at  $2\text{--}3$  °C. By gently blowing the water surface above the mounted slide with air supplied by an aquarium pump, we generated a stable low current velocity of approximately  $0.28 \pm 0.02$  (SE)  $\text{mm s}^{-1}$  in our observational area. We measured pH from a point close to the leaf surface up until out of the diffusive boundary layer, where the pH was stable. We used UNISENSE micro-pH sensors with 25 or 50  $\mu\text{m}$  tips, connected to a volt meter with 1-decimal precision for millivolt measurements (Consort, R362). pH sensors were calibrated with a three-point calibration using NIST buffers of  $\text{pH}_{\text{NBS}}$  4.0, 7.0 and 10.0 before each series of measurements. After each change in species or replica, a resting period of  $> 15$  min was observed to allow the diffusive boundary layer to be fully developed before measurements. A USB microscope (DinoCapture) connected to a PC with on-screen visualization software aided in visually establishing the lowest point of the measurements, as close to the macrophyte surface as possible without breaking the tip of the electrode. A scaled picture from this lowest point allowed for back-calculating the actual distance to the leaf surface afterwards. We allowed readings at this lowest point to stabilize for  $> 5$  min after which the millivolt value was written down manually. The microsensor was then raised 20  $\mu\text{m}$  with a precise 1-D micromanipulator and thereafter 30  $\mu\text{m}$ , after which we continued with 50  $\mu\text{m}$  increments and then 100 and 500  $\mu\text{m}$  increments until a stable pH was obtained for three measurements or more and we considered we were outside the diffusive boundary layer; between subsequent points the sensor was allowed to stabilize for at least 5 min. We evaluated 3 replicas of each species at a irradiance of  $200 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$ , and calculated the  $\Delta$  pH across the diffusive boundary layer (defined from the tissue surface to where pH was at  $0.99 \times$  water-column pH).

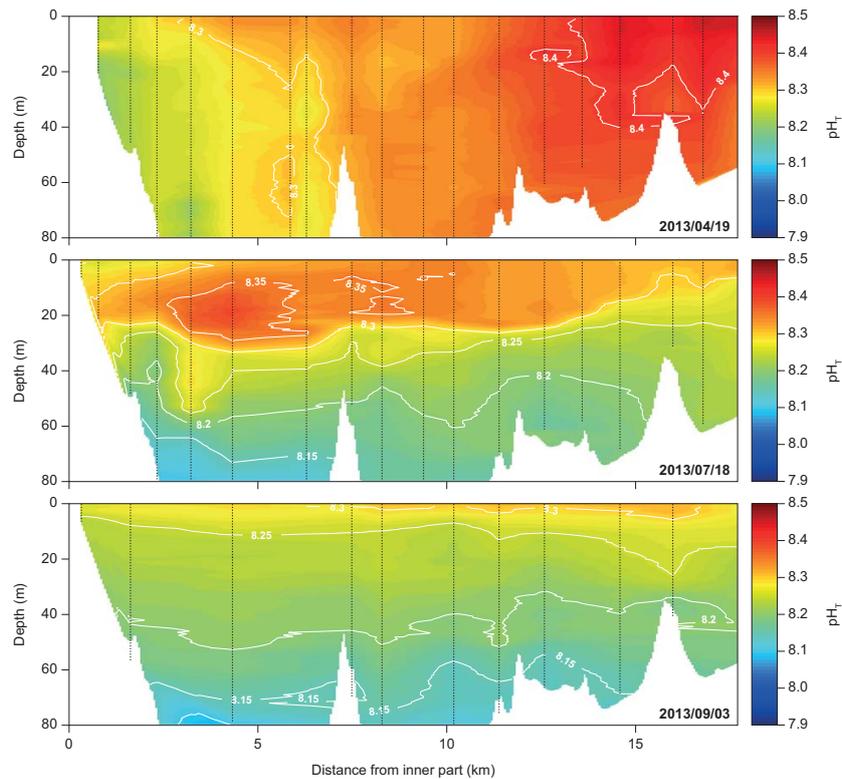
## 3 Results

Data are available in digital form (Krause-Jensen et al., 2015).

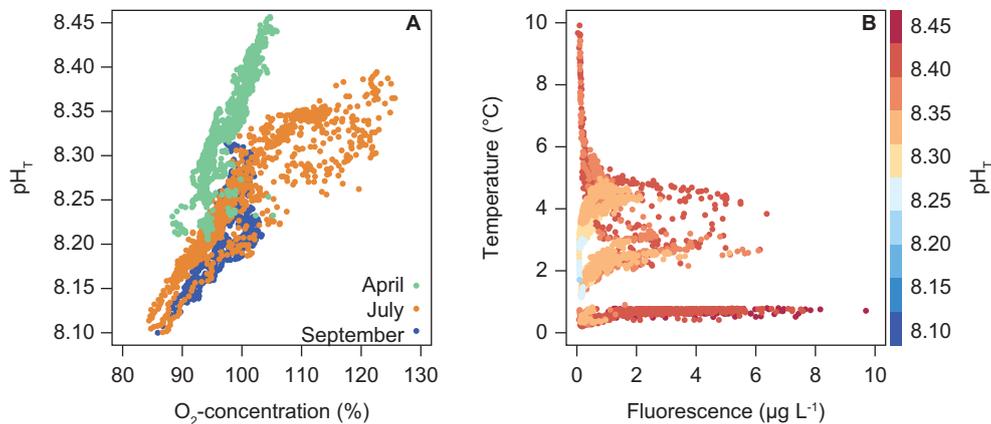
### 3.1 Fjord-scale and seasonal pH variability

Large seasonal and spatial variability was observed in pH values along the longitudinal gradient centrally in the fjord (Fig. 2a).  $\text{pH}_{\text{T}}$  in surface water increased in April due to  $\text{CO}_2$  consumption by the spring bloom as evidenced by a very high fluorescence (Fig. A2), to a maximum value of almost 8.50, most pronounced in the mouth of the fjord with values of around 8.25 in the inner part (Fig. 2). Accordingly, a horizontal gradient of around 0.25 pH units was observed along the main axis of the fjord.  $\text{pH}_{\text{T}}$  values in upper layers decreased during the summer to around 8.35 in July and with the maximum observed towards the inner part of the fjord. A further decrease in pH was observed in September, with more homogenous values in surface waters along the fjord gradient resulting in a horizontal range of only 0.05 pH units. Vertical gradients in pH from the surface to the deeper waters of the fjord ranged from only 0.1 units in April, when the fjord was vertically mixed, to 0.15 units in September to 0.25 pH units in July, when maximum  $\text{pH}_{\text{T}}$  values of 8.35 occurred in a subsurface algal bloom in the inner parts of the fjord with waters supersaturated in oxygen (up to 120 % saturation, Figs. A2, A3) and minimum values of  $\text{pH}_{\text{T}}$  8.1 were measured in the deeper sectors (Fig. 2a). Seasonally pH varied between 0.2 and 0.3 units in both surface and deep waters over the 5 months.  $\Omega_{\text{arag}}$  values were closely coupled to pH and ranged from minimum values of 1.6, observed in the bottom waters of the inner part of the fjord, to maximum values of 2.5 in the subsurface waters in September (Krause-Jensen et al., 2015). Corresponding  $p\text{CO}_2$  levels ranged from 162 to 325  $\mu\text{atm}$ , in the range of values recently reported for the fjord (Sejr et al., 2014).

Oxygen saturation at the fjord-scale ranged greatly from 85 to 127 % and was strongly related to pH for each of the three periods (Fig. 3a), pointing at strong biological control of pH variability within the fjord. The slope of the pH versus  $\text{O}_2$  relationship was steepest for the April survey, when the highest pH levels were observed. Examination of pH values in relation to fluorescence and temperature also showed that the warmest waters, of up to 10 °C, observed in July, supported intermediate pH, while the highest pH was observed in the coldest waters, corresponding to the April survey, when temperatures were uniformly low across the fjord (Fig. 3b). On a vertical scale, the cold bottom waters with low fluorescence generally supported the lowest pH values across seasons. Hence, overall, pH showed much tighter correlation with  $\text{O}_2$  levels than with water temperature, and the correlation between pH and  $\text{O}_2$  implied a similar close correlation between  $\Omega_{\text{arag}}$  and  $\text{O}_2$  levels.



**Figure 2.** Fjord-scale pH variability in Kobbefjord on 19 April, 18 July and 3 September 2013.



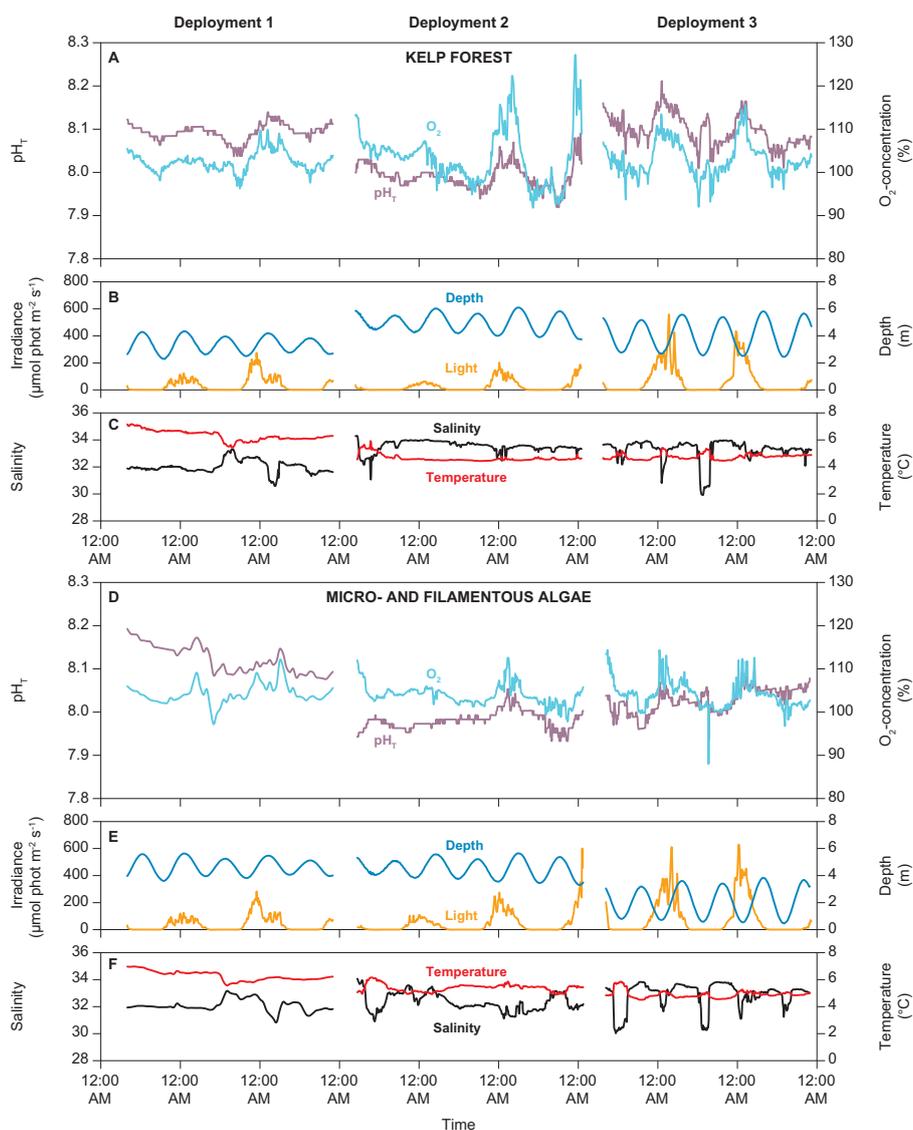
**Figure 3.** Fjord-scale relationships in Kobbefjord between pH and oxygen (a), and between temperature and fluorescence with associated pH levels shown with symbol colour (b), on three sampling occasions: 19 April, 18 July and 3 September 2013.

### 3.2 Small-scale and diurnal pH variability in kelp forests and benthic habitats colonized by microalgae/scattered filamentous algae

The three parallel deployments in kelp forest and habitats colonized by microalgae and scattered filamentous algae encompassed six complete diurnal cycles which exhibited peak  $\text{pH}_T$  levels during the day of 8.11 (8.04–8.19) (avg. (SD)) and 8.08 (8.02–8.16), respectively, as opposed to minimum

$\text{pH}_T$  levels during night of 8.02 (7.97–8.06) and 8.01 (7.94–8.09), respectively, with no significant difference between habitats ( $t$  test,  $p > 0.05$ ). The diurnal range of minimum night pH to maximum day pH was slightly higher in the kelp forest (avg.  $\pm$  SD =  $0.098 \pm 0.061$ ) than above the microalgae/filamentous algae ( $0.073 \pm 0.052$ ) (paired, one-tailed  $t$  test,  $p = 0.041$ ).

There were large differences in the extent of diel fluctuations in pH among deployments dependent on incident irra-

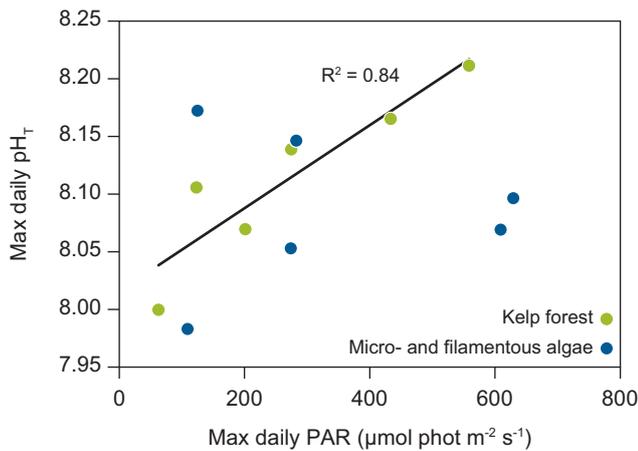


**Figure 4.** Diurnal variability in pH, O<sub>2</sub>, water depth (all measured by Hydrolab) and irradiance (measured by Odyssey loggers) at ca. 50 cm above the seafloor in kelp forests (a–c) and habitats colonized by microalgae/filamentous algae (e–f) during three parallel deployment in Kobbefjord, Nuuk, 27–30 August, 30 August–2 September, and 2–5 September 2013. The deployments represent the benthic sites (#1–3, respectively) shown on the map (Fig. 1).

diance and the shifting phase of tidal state and the solar cycle (Fig. 4). Diel pH fluctuations were small during dark, cloudy days and when high tide coincided with peak solar radiation, thereby reducing incident irradiance on the benthic habitat. In contrast, diel pH fluctuations were amplified in deployments during sunny days when low tide coincided with peak solar radiation (Fig. 4). Hence, the interaction between tide and the solar cycle controlled incident radiation and thereby induced fluctuations in photosynthetic activity and pH. This was particularly apparent in kelp forests where peak daily pH increased as a function of maximum daily photosynthetic solar radiation reaching the habitat during the day, whereas this relationship was not significant in the water column above

the microalgae/filamentous algae (Fig. 5). Indeed, biologic control of pH was also reflected in strong relationships between pH and O<sub>2</sub> concentration within each deployment in the kelp forests ( $R^2 = 0.64\text{--}0.76$ ), particularly during high irradiance, as opposed to weaker pH versus O<sub>2</sub> relationships for the microalgae/filamentous algae sites ( $R^2 = 0.05\text{--}0.15$ ) which also showed much smaller variability in O<sub>2</sub> levels (98–114 % saturation) than did the kelp forest (92–128 % saturation) (Fig. 6). The diurnal range of O<sub>2</sub> concentrations in the kelp forest matched the range recorded at pelagic fjord scale on a seasonal basis (85–127 %, Fig. 3).

Tidal changes in water masses, reflected by changes in salinity and temperature, also contributed to variations in



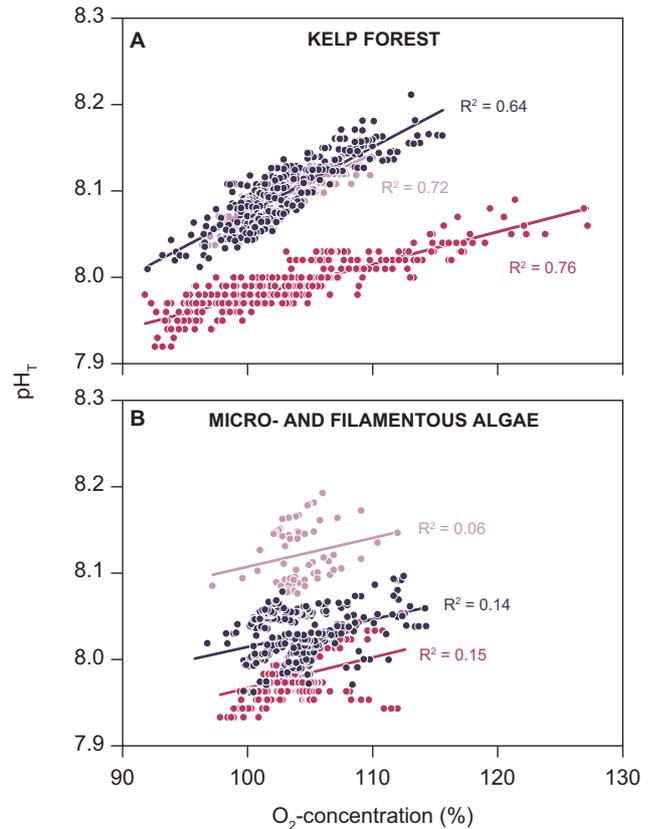
**Figure 5.** Maximum daily pH in a kelp forest (green dots) and above microalgae/filamentous algae (blue dots) as a function of maximum daily incident light over 6 full days as measured during three parallel deployments in Kobbefjord, Nuuk, 27–30 August, 30 August–2 September, and 2–5 September 2013. Linear fit and coefficient of determination are shown for the significant relationship for the kelp forest.

pH and  $O_2$  levels. This was visible as incidences of sudden changes in pH paralleling fluctuations in salinity and also as differences in pH levels between deployments in water masses of different salinity (Fig. 4). However, salinity explained much less of the variation in pH than did  $O_2$ , except in one deployment in the microalgae/filamentous algae habitat when salinity explained 51 % of the variation in pH as opposed to 15 % explained by  $O_2$  ( $R^2 = 0.04$ – $0.33$  in kelp forest;  $R^2 = 0.04$ – $0.51$  in microalgae/filamentous algae, data not shown). Thus, overall biological activity had a much stronger influence on pH than had exchange of water masses.

The observed diurnal pH variability also translated into important fluctuations in  $\Omega_{\text{arag}}$ , involving  $0.18 \pm 0.06$  units (from maximum day levels of  $1.77 \pm 0.21$  to minimum night levels of  $1.60 \pm 0.17$ ) in the kelp forest and  $0.14 \pm 0.07$   $\Omega_{\text{arag}}$  units (from maximum day levels of  $1.72 \pm 0.30$  to minimum night levels of  $1.58 \pm 0.26$ ) at the microalgae/filamentous algae sites. Corresponding  $p\text{CO}_2$  levels ranged from 238 to 536  $\mu\text{atm}$  at the kelp sites and from 258 to 515  $\mu\text{atm}$  at the microalgal/filamentous algal sites.

### 3.3 Metre- to millimetre-scale pH variability in kelp forests

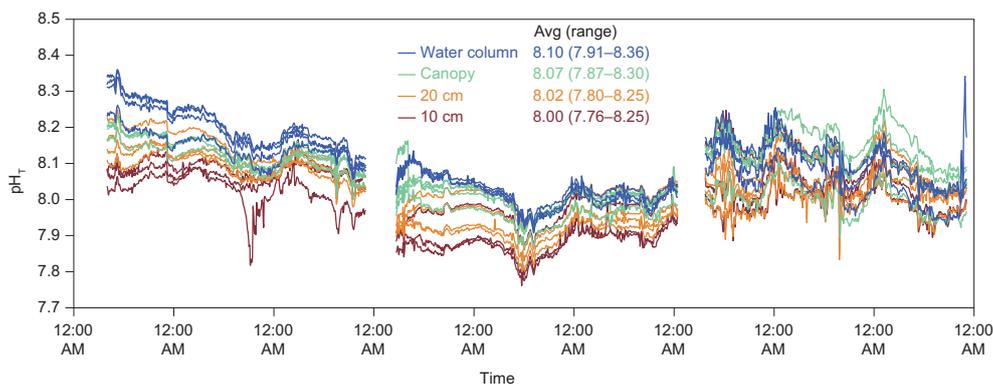
Examination of the variability in pH within  $1 \text{ m}^3$  kelp forest, sampled from the bottom of the canopy to the overlying water column, using the multi-electrode array, showed very large concurrent pH variability involving about 0.2 to 0.3 pH unit differences at any given time and with a total  $\text{pH}_T$  range of 7.76–8.36 across deployments (Fig. 7). In general, pH tended to be highest at the top of the canopy and in the water just above the canopy, reflecting that the canopy top is the



**Figure 6.** pH vs.  $O_2$  concentration for three parallel deployments (#1–3 shown by increasing colour intensity) in subtidal habitats colonized by kelp forests (top panel) or microalgae/scattered filamentous algae (bottom panels) in Kobbefjord, Nuuk, August–September 2013. Each deployment represents 10 min loggings by multiloggers (Hydrolab) over ca. two diurnal cycles. Linear fits and coefficients of determination are shown.

most photosynthetically active layer, while pH was generally lower in the shaded bottom part of the canopy (Fig. 7), where photosynthetic biomass and incident irradiance are lower and respiration rates higher. The range of pH within  $1 \text{ m}^3$  of kelp forest at any one point in time was comparable among deployments, despite the different light conditions, although the absolute values of pH differed among deployments, with highest levels observed at peak incident irradiance (Fig. 7). This small-scale variability in pH also translated into a variability in  $\Omega_{\text{arag}}$  of about 0.20 units in  $1 \text{ m}^3$  of habitat at any time.

pH also varied significantly within the diffusive boundary layer of the six macrophyte species examined in the light (Fig. 8a), with pH increasing by 0.07–0.85 units, depending on species, from the top of the 0.3–2.2 mm thick diffusive boundary layer to the surface of the plants (Fig. 8b).



**Figure 7.** pH variability within  $1\text{ m}^3$  of kelp forest in Kobbefjord, Nuuk, during three deployments in late August–September 2013. Sixteen pH sensors were configured in situ in a 3-day array with four sensors at 0.1 m from the bottom, four sensors at 0.2 m, four sensors just underneath the canopy and four in the water column above the canopy, which typically extended about 0.75 m above the seafloor.

### 3.4 pH variability in intertidal pools

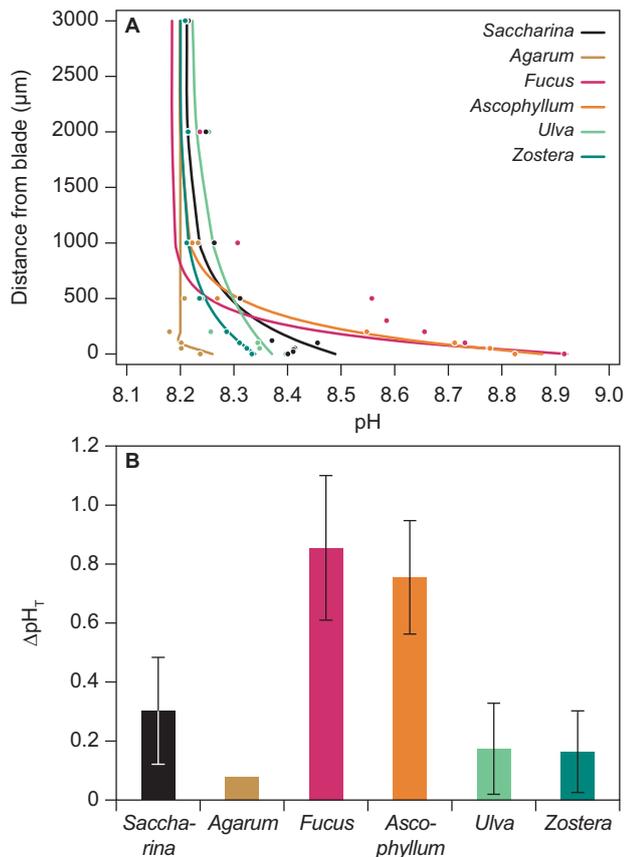
pH and oxygen concentration showed important diel variability in vegetated intertidal pools, with oxygen supersaturation (up to 176 %) during the day and undersaturation (down to 11 %) at night, compared to far more uniform concentrations in the surface waters on the adjacent vegetated shore (89–111 % saturation, Fig. 9). Accordingly,  $\text{pH}_T$  changed greatly in intertidal pools, reaching maximum values of 9.0 during the day and minimum values of 7.4 during night periods, i.e. a diel range of ca. 1.6 pH units. Diel pH fluctuations in the surface waters of the adjacent shore were much smaller (8.0–8.5) but still high, reflecting the metabolic activity of the intertidal vegetation growing on the shore (Fig. 9). The difference in pH between vegetated intertidal pools and adjacent shores provided an additional example of variability in pH between adjacent habitats.

## 4 Discussion

Our results highlight the nested scales of variability in pH present in the Kobbefjord ecosystem involving (1) seasonal variability, largely driven by the phytoplankton spring bloom as a major event affecting pH; (2) diel variability acting through complex changes in submarine irradiance modulating rates of photosynthesis and respiration of benthic vegetation driven by the interaction of the solar and the tidal cycles; (3) large-scale variability along horizontal and vertical fjord gradients reflecting gradients in metabolic activity in combination with movement of water masses; (4) variability between subtidal habitats with and without kelp forests and between vegetated tidal pools and adjacent vegetated shores reflecting variable degrees of biological control; (5) small-scale three-dimensional variability due to heterogeneity in metabolic processes and mixing in vegetated habitats; and (6) microscale variability across the diffusive boundary layer of macrophytes (Fig. 10).

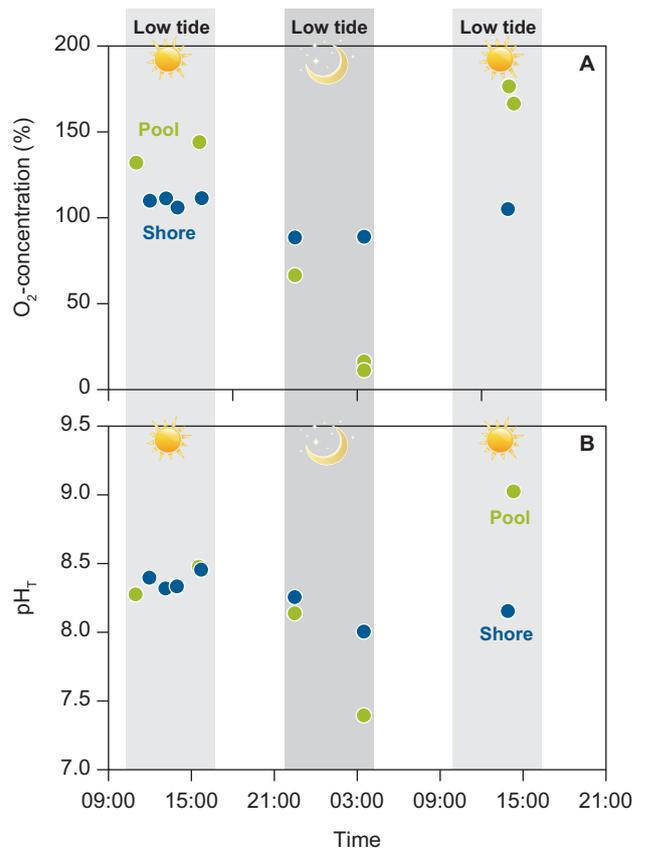
Overall, metabolic processes played a fundamental role in driving pH variability across scales, as reflected in strong relationships between oxygen concentration and pH at the fjord scale and at both diel and seasonal scales. Primary producers played a major role in the regulation of pH variability, both in the pelagic zone, where in particular the intense spring bloom characteristic of Arctic ecosystems (Takahashi et al., 2003; Sejr et al., 2014) induced high pH in the subsurface layers while the respiratory process in the bottom waters reduced pH, and in the nearshore benthic environment, where the presence of subtidal kelp forests and intertidal macroalgae induced marked spatial and diurnal variability in pH. The mosaics of pH reflected that the density of the primary producers, and the spatio-temporal separation of photosynthesis and ecosystem respiration in combination with mixing of water masses were key drivers of the variability in both planktonic and benthic communities. Hence, the vertical gradient of declining pH from upper illuminated to lower shaded habitats varied from the 10–100 m scale in the planktonic community where the density of primary producers is relatively low to the centimetre to metre scale in dense kelp forests. The same is true on a temporal scale where the diurnal pH variation in the benthic vegetation matches the seasonal variability in pH in the planktonic community.

The scale of seasonal pH variability in the planktonic community (Fig. 10) compared well with previous reports for the Arctic, showing the spring bloom as a prevalent driver of  $p\text{CO}_2$  (Sejr et al., 2011; Meire et al., 2015). Though a multitude of factors including water depth, light regime, season, seawater retention time, density and plant species may all affect pH variability in vegetated habitats, our results match evidence from other latitudes of strong pH variability in macroalgal forests and seagrass meadows. Hence, marked diel pH variability has also been reported from a Californian kelp forest (Frieder et al., 2012), from a Mediterranean seagrass bed (Hendriks et al., 2014), and, in an extreme case, from a temperate, shallow, dense algal bed (diel range: ca. 1



**Figure 8.** Microscale pH variability across diffusive boundary layers of blades of six different macrophyte species illuminated by  $200 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ : the kelps *Saccharina longicuris* and *Agarum clathratum*, the intertidal brown macroalgae *Fucus vesiculosus* and *Ascophyllum nodosum*, the green macroalga *Ulva lactuca*, and the seagrass *Zostera marina*. (a) pH levels (mean of 2–3 replicate measurements) across blade diffusive boundary layers fitted by an exponential model ( $y = y_0 + a \times \exp^{-b \times x}$ ,  $R^2 > 0.90$  for all individual fits). (b) pH range across the diffusive boundary layer of the various species.

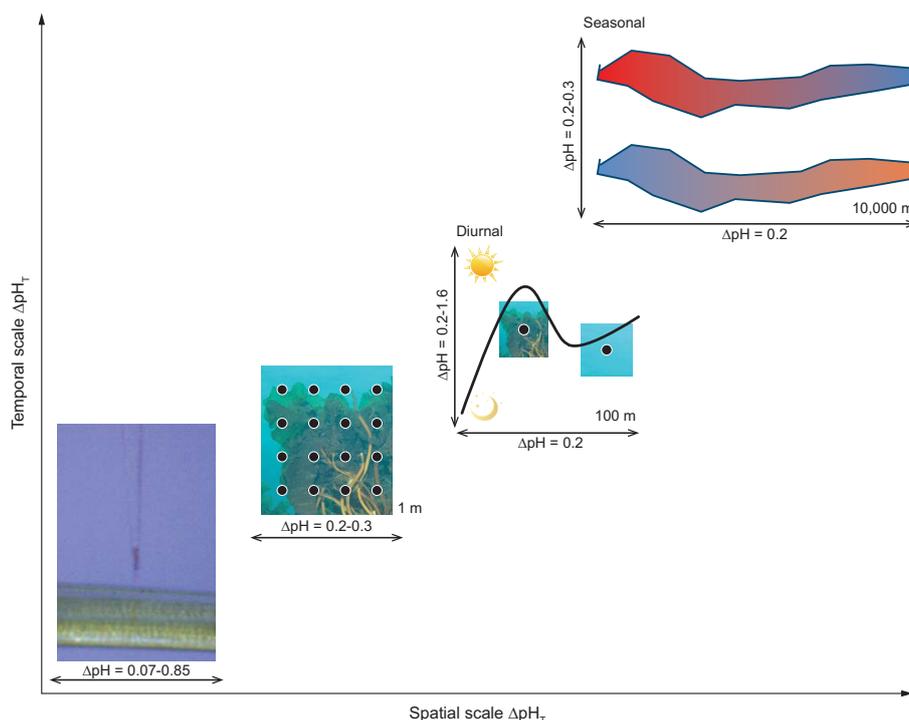
unit; Middelboe and Hansen, 2007) and kelp forest (diel range: ca. 0.6–0.8 pH units; Cornwall et al., 2013a). Our pH measurements in benthic habitats neighbouring the kelp forest also carried a biological signal, though less distinct, likely reflecting the combined signal of the benthic primary producers at the site, of the neighbouring kelp forests, and of the planktonic community in the water masses exchanged with tidal currents. The marked biological control of pH in kelp forests suggests that diel pH may be even more pronounced during sunny days with more intense photosynthesis than during the generally overcast conditions of our survey. Thus, while the identified pH range and pH vs.  $\text{O}_2$  relationships for the planktonic community covered the full growth season, they solely represented a few overcast September days in the benthic habitats and would likely involve markedly



**Figure 9.**  $\text{O}_2$  concentration and pH in vegetated tidal pools and in surface waters of neighbouring vegetated intertidal shores measured at low tide during day and night just after pool formation and before pool inundation.

higher levels had they covered the full growth season. For subantarctic giant kelp forests, the diel amplitude in  $p\text{CO}_2$  and  $C_T$  (Delille et al., 2009) during spring and summer, as well as the seasonal amplitude in pH,  $C_T$  and  $p\text{CO}_2$  (Delille et al., 2000), was reported to be markedly higher within kelp forests as compared with unvegetated habitats, underlining the kelps' strong biological control of pH.

We further show, for the first time, significant 3-D variability in pH within  $1 \text{ m}^3$  of kelp forest, with pH ranging about 0.2–0.3 pH units at any one point in time and a total variability across deployments of 7.76–8.36  $\text{pH}_T$ , resembling the range recorded across the entire growth season in the pelagic. Levels of pH were dependent on the position in the kelp canopy, with the highest pH generally appearing at the top of the canopy and decreasing toward the seafloor, likely reflecting the vertical structure of photosynthetic activity in the kelp bed. The fast rates of metabolic activity in combination with reduced flow in such densely vegetated habitats make these 3-D patterns appear in spite of the marked exchange of water masses resulting from the 1–4.5 m tidal range.



**Figure 10.** Conceptual summary of nested scales of temporal and spatial variability in pH in Kobbefjord, Nuuk. The figure shows the maximum pH range at the various scales examined. From lower left to upper right: (1) microscale variability across macrophyte diffusive boundary layers, (2) small-scale variability within kelp forests, (3) diurnal variability in vegetated subtidal habitats and intertidal pools/adjacent shores and variability between habitats at the 100 m scale, and (4) seasonal and fjord-scale horizontal variability.

Changes in pH were particularly pronounced in small tidal pools, where photosynthesis of dense seaweed stands of primarily *Ascophyllum nodosum* and *Fucus* spp. drove  $O_2$  levels to large supersaturation levels (176 %) and forced pH to extremes of up to  $pH_T$  9.0 at low tide during sunny days, corresponding to  $\Omega_{arag}$  of 4.14 and  $pCO_2$  of 13  $\mu atm$  compared to night values of  $pH_T$  7.4,  $\Omega_{arag}$  of 0.27 and  $pCO_2$  of 1647  $\mu atm$  driven by community respiration, which almost depleted  $O_2$  in the pools (11 % saturation). In surface waters of adjacent densely vegetated intertidal shores, we observed a maximum  $pH_T$  of 8.5 with corresponding  $\Omega_{arag}$  2.23 and  $pCO_2$  of 96  $\mu atm$  during the day and a minimum  $pH_T$  of 8.0, with corresponding  $\Omega_{arag}$  of 0.54 and  $pCO_2$  of 243  $\mu atm$  during the night. While intertidal brown macroalgae thrive in such habitats when regularly flushed as in the current study, apparently only *Ulva* (*Enteromorpha*) *intestinalis* occurs in isolated, rarely flushed rock pools, where it can drive pH to levels > 10 (Björk et al., 2004).

At the microscale, pH also showed considerable variability with a range of up to 0.85 pH units across the diffusive boundary layer of the key species of the vegetated shallow ecosystems, with high pH levels at the tissue surface declining towards the bulk water during daytime (Fig. 8). There was substantial variability among species, with intertidal macroalgae (*Ascophyllum* and *Fucus*) showing the

largest pH range. The interspecific differences likely related to the species' photosynthetic rates as well as to their morphology, which affect the thickness of the diffusive boundary layer (Hurd and Pilditch, 2011). This microscale pH variability across the diffusive boundary layer compared well with previous observations for the calcifying alga *Halimeda discoidea* (pH range of 0.7 across diffusive boundary layer; de Beer and Larkum, 2001) as well as for the coralline alga *Sporolithon durum* (light–dark pH change at tissue surface 0.9; Hurd et al., 2011) and *Arthrocardia corymbosa* (pH range across diffusive boundary layer 0.4, for example, depending on flow; Cornwall et al., 2013b). The pH range across the diffusive boundary layer of *Ulva* was surprisingly low considering the ability of *Ulva* to elevate pH to high levels (Björk et al., 2004), but it was probably the combination of low water temperature and limited nutrient supply that limited the photosynthetic rate. The diffusive boundary layer thickness as well as the pH range across it depends markedly on flow conditions. Reduced flows as present in dense vegetation increase the diffusive boundary layer thickness and consequently the pH range (Hurd et al., 2011; Cornwall et al., 2013b). The current experiment was, hence, conducted at reduced flow and, importantly, at the same flow for all species. Exchange of water masses with different salinity and temperature also added to the variability in pH as indicated for both

pelagic (Fig. 3b) and benthic (Fig. 4) systems but showed much weaker correlation to pH than did  $O_2$  concentrations reflecting the biological control.

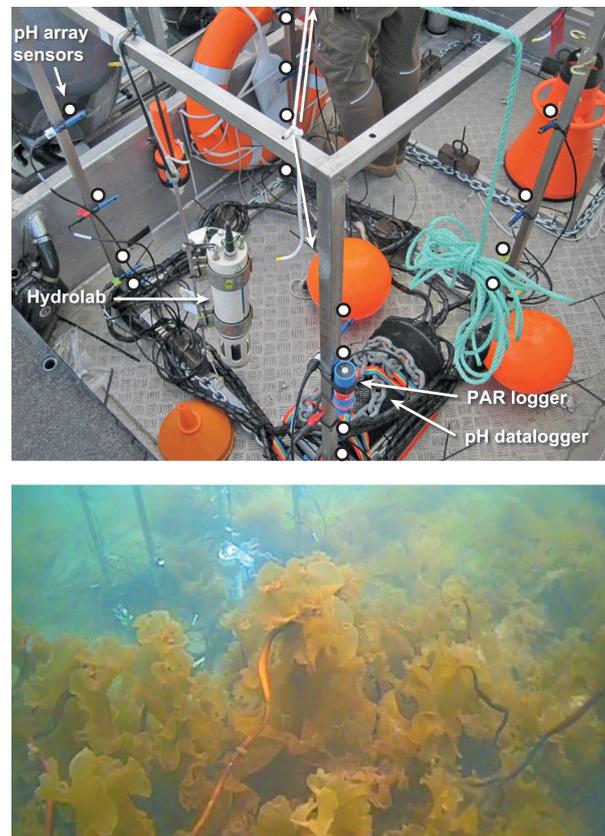
The processes above resulted in nested scales of pH variability in the Kobbefjord ecosystem (Fig. 10), with variability ranging 0.2–0.85 units across spatial scales and 0.2–1.6 units over diurnal to seasonal scales. This variability provides a dynamic mosaic of niches for organisms. Niches of high pH may be particularly important for the more vulnerable larval and juveniles stages of calcifiers under conditions of low pH as projected for the future (Kroecker et al., 2013). The suitability for calcifiers is best represented by  $\Omega_{\text{arag}}$ , where calcifiers should be favoured by high  $\Omega_{\text{arag}}$  values. The Kobbefjord ecosystems host a number of calcifying species, including bivalves such as blue mussels, scallops and snails; echinoderms such as green sea urchins; crustaceans such as *Pseudobalanus balanoides*; and calcareous algae and foraminifers. Overall, the identified  $\Omega_{\text{arag}}$  conditions were well above 1, particularly in illuminated habitats with intense photosynthesis and, hence, indicated favourable conditions for calcification. The phytoplankton spring bloom, depleting  $CO_2$  and driving  $\Omega_{\text{arag}}$  to values close to 3, would also provide adequate conditions for pelagic calcifiers, as it would provide the double benefit of adequate environments for aragonite deposition and food supply to support growth and the energetic demands of calcifiers. Canopies of kelp and intertidal seaweed environments may also provide adequate niches for calcifiers during summer, when  $\Omega_{\text{arag}}$  values would be highest through the cumulative action of the processes upregulating pH and  $\Omega_{\text{arag}}$  values discussed above. Indeed, most calcifiers spawn and recruit in early summer (Arendt et al., 2013), when  $pCO_2$  remains low, warmer water temperatures lead to higher  $\Omega_{\text{arag}}$  and high solar radiation and a long photoperiod allow seaweeds to draw down  $CO_2$  further (Delille et al., 2000).

The upregulating effect of primary producers on pH is counterbalanced by the opposite effect of respiration and decomposition prevailing in shaded and deeper basins and periods as illustrated by the large-scale seasonal variability in the pelagic community (Fig. 2), and paralleled in kelp forests outside the productive period (Delille et al., 2009) as well as during night-time and in shaded layers of the kelp forest (Fig. 7) and tidal pools (Fig. 9). These shaded habitats with diurnally low  $\Omega_{\text{arag}}$  could be challenging habitats for calcifiers. Interestingly, however, blue mussels grew in close association with macroalgae even in intertidal pools, where they would experience maximum  $\Omega_{\text{arag}}$  values of up to 4.28 when low tide occurred at noon as opposed to levels as low as 0.28 during night (Fig. 9). Blue mussels have indeed been observed to abound in intertidal macroalgal habitats (Blicher et al., 2013) and along with other calcifiers to be trophically linked with habitat-forming algae such as *Ascophyllum* (Riera et al., 2009), and have also been reported to tolerate high  $pCO_2$  concentrations when food is abundant (Thomsen et al., 2013). The recurring periods of high  $\Omega_{\text{arag}}$

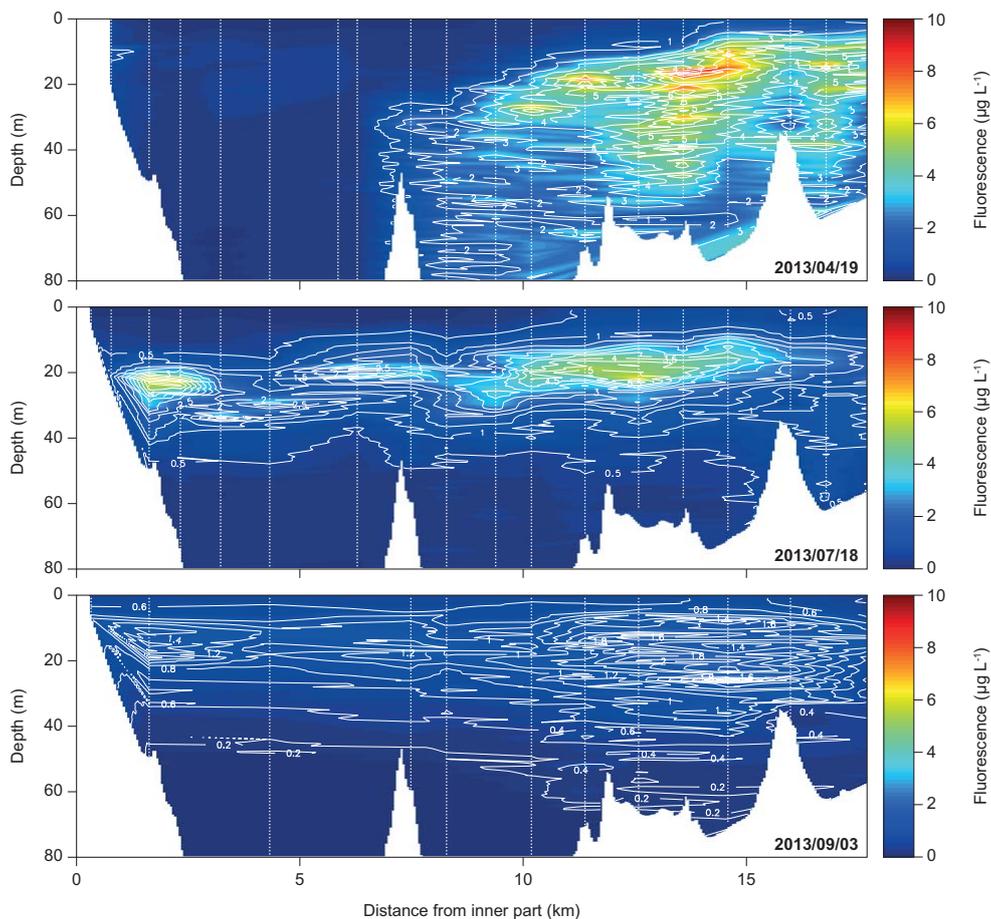
in combination with adequate food supply can likely compensate for the potential problems of low  $\Omega_{\text{arag}}$  during night. Laboratory experiments have demonstrated that semidiurnal fluctuations of 0.3 pH units may compensate for negative effects of constantly low pH on the development of mussel larvae (Frieder et al., 2014). Calcareous epiphytic organisms, such as encrusted algae and bryozoans, would also experience high variability in  $\Omega_{\text{arag}}$  at the surface of the plant tissue, where periodically high  $\Omega_{\text{arag}}$  values favours calcification, as elegantly demonstrated by de Beer and Larkum (2001).

The existence of a mosaic of environments in the Kobbefjord underlines the importance of metabolic processes along with habitat configuration and interactions among community constituents in affecting pH in coastal ecosystems as opposed to the simpler situation in the open ocean (Duarte et al., 2013; Hendriks et al., 2014). This pronounced influence of metabolic processes occurs in spite of Kobbefjord being a macrotidal area with marked exchange of water masses with the coastal region and is probably also the case in many other shallow coastal areas in the Arctic, as has also been highlighted for areas in the temperate zone (Duarte et al., 2013). While the current study explored pH in benthic habitats under overcast situations in the early autumn of the subarctic, kelp forests are likely to induce much more pronounced increases in pH and  $\Omega_{\text{arag}}$  in midsummer, when irradiances are higher and the photoperiod longer, and further north, during high-Arctic midsummer, when the sun does not set for months. Under scenarios of ocean acidification such vegetated habitats may gain increased importance as local refuges for calcifiers. The projected poleward expansion of macrophytes into the Arctic with warming and reduced sea-ice cover (Müller et al., 2009; Jueterbock et al., 2013) has been hypothesized to provide such niches of elevated pH and  $\Omega_{\text{arag}}$  during summer (Krause-Jensen et al., 2014). Similarly, increased pelagic primary production as forecasted for parts of the Arctic Ocean (Arrigo et al., 2008; Slagstad et al., 2011; Popova et al., 2012) may also create local niches of high pH.

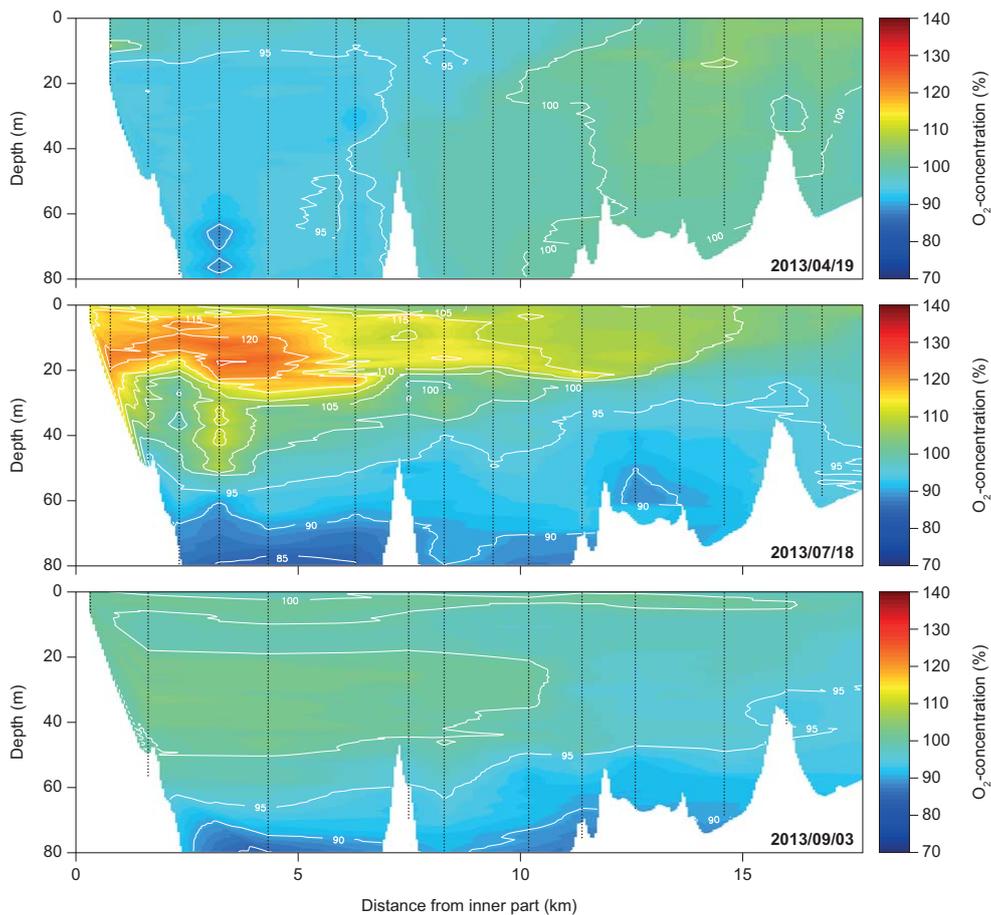
## Appendix A



**Figure A1.** Photo of deployment frame with loggers shown on the deck of the boat (upper panel) and in situ in the *Saccharina longicruris*-dominated kelp forest (site #1, central panel). Markings in upper panel show the array of 16 pH sensors connected to a common pH logger; the Hydrolab instrument measuring salinity, temperature and oxygen; and a PAR logger (Odyssey).



**Figure A2.** Fjord-scale variability in fluorescence in Kobbefjord, Nuuk, 19 April, 18 July and 3 September 2013.



**Figure A3.** Fjord-scale variability in O<sub>2</sub> concentration in Kobbefjord, Nuuk, 19 April, 18 July and 3 September 2013.

*Author contributions.* Planning, field work, data processing and writing were carried out jointly, led by D. Krause-Jensen and C. M. Duarte, with L. Meire and M. K. Sejr responsible for fjord-scale surveys; I. E. Hendriks, M. K. Sejr, M. E. Blicher, C. M. Duarte and D. Krause-Jensen responsible for the various small-scale measurements; I. E. Hendriks and N. Marba for microscale measurements; N. Marba and D. Krause-Jensen for intertidal measurements; and M. E. Blicher for Greenland field facilities. Main idea: C. M. Duarte.

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